

Poster

AnABlast: a useful tool to analyse the *Caenorhabditis elegans* genome sequence

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ABSTRACT

Motivation: During the last decade, genomes analysis has experienced great advances. However, genomes still hide mysteries to be decoded. There are many computer programs that can assist us to better understand genomes and to their annotation. AnABlast (Ancestral-patterns search through A BLAST-based strategy) was developed to detect small and complex hidden coding sequences. It generates profiles of alignments on query amino acid sequences using lo-stringency BLAST strategy. Three years ago, AnABlast was tested in a genome-wide search for coding sequences in *Schizosaccharomyces pombe* and several new genes and ancient coding sequences were efficiently identified in the fission yeast genome. Due to this huge success, this strategy is now being applied to *Caenorhabditis elegans*. In this work, we have used AnABlast to search the *C. elegans* genome for new, uncharacterized putative coding sequences. As a functional assay we have performed, RNA interference (RNAi) against these sequences to check if a visible phenotype could be observed. Upon genome analysis AnABlast hits show up as the alignments along query sequences accumulation peaks in possible coding regions. Although, the genome of *C. elegans* is well annotated it could still be challenging because AnABlast has discovered several peaks that could probably be new genes or undescribed exons from previously of annotated genes.

Methods: The methodology consists in two main processes. First, we searched for possible new coding sequences by AnABlast. Second, we are constructing a library to perform RNAi experiments against all possible sequences chosen. In order to do that, primers were generated to amplify each selected sequence. The resulting fragments are cloned on the RNAi producing plasmid L4440 and transformed into the *Escherichia coli* HT115 strain, resulting in individual bacterial strains, each designed to produce RNAi against a single *C. elegans* predicted peak. These *E. coli* expressing the dsRNA are then used to perform separate RNAi experiments to individually analyze the possible phenotype of each selected nucleotides sequences.

Conclusions: With this, we will have a very potent RNAi library of possible new genes that could be very useful in the future. Moreover, we are demonstrating that AnABlast is a potent tool to uncover new genes, and it may be applicable to the "in silico" analysis of others organism sequences genome.

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